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for

**“METHOD FOR FABRICATING A BIOCHIP USING THE HIGH DENSITY CARBON
NANOTUBE FILM OR PATTERN”**

Cross-Reference to Related Application:

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METHOD FOR FABRICATING A BIOCHIP USING THE HIGH DENSITY CARBON NANOTUBE FILM OR PATTERN

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BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates to a biochip comprising a bio-receptor chemically
10 or physicochemically attached to a high density carbon nanotube (CNT) film or pattern
having chemical functional groups exposed, in which the bio-receptor is capable of
binding to a target biomaterial, and a method for preparing the same.

Background of the Related Art

15 Carbon nanotube (CNT) is an allotrope of carbon, which consists of carbons
exists abundantly on the earth. They are tubular materials where a carbon atom is
connected to other carbons in the form of a hexagonal honeycomb structure. Their
diameter is about size of nanometer ($1/10^9$ meter). CNT is known to have excellent
mechanical properties, electrical selectivity, field emission properties and highly
20 efficient hydrogen storage properties and be new and almost defect-free of all the
existing materials.

Because of their properties of excellent structural rigidity, chemical stability,
ability to act as ideal one-dimensional (1D) "quantum wires" with either
semiconducting or metallic behaviors and a large aspect ratio, CNT exhibits a broad
25 range of potential applications as a basic material of flat panel displays, transistors,

energy reservoirs, etc., and as various sensors with nanosize (Dai, H., *Acc. Chem. Res.*, 35:1035-1044, 2002).

In order to apply such properties more diversely, the purified single-walled CNT has been cut into short nanotube pieces using an acid. The cut CNT pieces have
5 mainly -COOH chemical functional groups at a part of ends and sidewall of the open tube. The properties of the CNT have been modified by chemical binding of various materials using these chemical functional groups. Further, there have been reported that the functional group of CNT was substituted for -SH group by chemical manipulation and patterned on a gold surface (Nan, X. *et al.*, *J. Colloid Interface Sci.*,
10 245:311-8, 2002) and that CNT was immobilized on substrate using the electrostatic method (Rouse, J.H. *et al.*, *Nano Lett.*, 3:59-62, 2003). However, the former has disadvantages of the low CNT surface density and the weak bonding, and the latter also has a fatal disadvantage that the patterning method for selective immobilization on the surface cannot be applied. Therefore, there is an urgent demand for developing a
15 new type surface immobilizing method with high density.

Recently, researches are being conducted to detect reactions both protein-protein and protein-ligand by means of electrochemical changes of CNT after immobilization of a biomaterial (Dai, H. *et al.*, *ACC. Chem. Res.*, 35:1035-44, 2002; Sotiropoulou, S. *et al.*, *Anal. Bioanal. Chem.*, 375:103-5, 2003; Erlanger, B.F. *et al.*,
20 *Nano Lett.*, 1:465-7, 2001; Azamian, B.R. *et al.*, *JACS*, 124:12664-5, 2002). A representative example of a protein-ligand reaction is an avidin-biotin reaction. Star *et al.* formed a channel on a substrate, which had been treated with a polymer, using CNT and measured the binding activity of streptoavidin by means of an electrochemical method (Star, A. *et al.*, *Nano Lett.*, 3:459-63, 2003).

25 The reasons that CNT attracts public attention as a biochip are as followings:

Firstly, it needs no labeling; secondly, it has high sensitivity to signal change; and thirdly, it is capable of reacting in an aqueous solution without deterioration of a protein. The combination of a new nanomaterial and a biological system will create important fusion technologies in respective fields of disease diagnosis (hereditary diseases), proteomics and nanobiotechnology.

In order to develop a rapider and cheaper biochip, many researches have been conducted on technologies of DNA hybridization detection. Various labeling technologies for detecting DNA hybridization have been developed. An effective surface treatment capable of increasing hybridization efficiency and simultaneously, removing the background from non-specific binding is required to detect the DNA hybridization effectively using the DNA chip. Many researches have been conducted to prepare a surface-treated DNA chip platform (Rogers, Y. *et al.*, *Anal. Biochem.*, 266:23-30, 1999; Hu, J. *et al.*, *Nuc. Acid. Res.*, 29:106-10, 2001). Also, various methods for detecting DNA hybridization were developed, which include the scanometric method, the colorimetric method, a method using nanoparticle, a method using electrochemistry, and etc. (Taton, T.A. *et al.*, *Science*, 289:1757-60, 2000; Alexandre, I. *et al.*, *Anal. Biochem.*, 295:1-8, 2001; Cai, H. *et al.*, *Analyst.*, 127:803-8, 2002; Cai, H. *et al.*, *Anal. Bioanal. Chem.*, 375:287-93, 2003).

Besides many applications with CNT in the bioengineering field have been recently being appeared. It is being suggested that the applicability of CNT to biochips, such as glucose biosensors, detecting protein, detecting a certain DNA sequence and the like (Sotiropoulou, S. *et al.*, *Anal. Bioanal. Chem.*, 375:103-5, 2003; Chen, R.J. *et al.*, *Proc. Natl. Acad. Sci. USA*, 100:4984-9, 2003; Cai, H. *et al.*, *Anal. Bioanal. Chem.*, 375:287-93, 2003). Screening bio-molecules from multilayer based on CNT can increase the amount of immobilized bio-substances, such as DNAs and detecting

sensitivity to the bio-substances, since the CNT has wide surface area and high electric conductivity.

At the present time, the most universal method for detecting the result of the reaction in a biochip is to use conventional fluorescent materials and isotopes (Toriba, A. *et al.*, *Biomed. Chromatogr.*, 17:126-32, 2003; Syrzycka, M. *et al.*, *Anal. Chim. Acta*, 484:1-14, 2003; Grow, A.E. *et al.*, *J. Microbio. Meth.*, 53:221-33, 2003). However, as novel methods to easily and precisely measure an electrical or electrochemical signal are attempted, there are increased demands for CNT as a new material.

The methods comprising preparing a high density CNT multiplayer, attaching DNA thereon and detecting complementary DNA are useful in genotyping, mutation detection, pathogen identification and the like. It has been reported that PNA (peptide nucleic acid: DNA mimic) is regio-specifically fixed on a single walled CNT and the complementary binding to probe DNA is detected (Williams, K.A. *et al.*, *Nature*, 420:761, 2001). Also, there has been an example, in which an oligonucleotide was fixed on a CNT array by a electrochemical method and DNA was detected by guanine oxidation (Li, J. *et al.*, *Nano Lett.*, 3:597-602, 2003). However, these methods do not apply CNT to fabrication and development of biochips.

Recently, a high capacity biomolecule detection sensor using CNT was disclosed (WO 03/016901 A1). This patent relates to a multi-channel type biochip produced by arranging a plurality of CNTs on a substrate using a chemical linker and attaching various types of receptors. However, it has a disadvantage of relative weakness to environmental changes.

Therefore, the present inventors have found a method for producing a CNT-biochip by repeated laminating CNT on a substrate having exposed amine groups by

chemical bonding to form a high density CNT film or pattern having exposed chemical functional groups and chemically binding a bio-receptor to the CNT film or pattern, or treating the CNT surface with a chemical to prevent the non-specific binding by physical adsorption and chemically binding the bio-receptor to the treated surface, and
5 completed the present invention.

SUMMARY OF THE INVENTION

10 It is an object of the present invention to provide a high density CNT film repeatedly immobilized on a substrate by chemical bonding and having chemical functional groups exposed on its surface, a CNT-biochip comprising bio-receptors attached onto the surface of the CNT film, and a method for preparing the same.

It is another object of the present invention to provide a CNT-biochip
15 comprising a bio-receptor attached to a desired position on a high density CNT pattern laminated by chemical bond and a method for preparing the same.

It is a further object of the present invention to provide a method for detecting various target biomaterials capable of binding to or reacting with a bio-receptor using the CNT-biochip.

20 To achieve the above objects, the present invention provides a method for producing a high density CNT film or pattern having a carboxyl group, exposed on its surface, which comprises the steps of: (a) reacting a substrate having amine groups exposed on the surface or a substrate having amine groups exposed in a patterned substrate with CNT having exposed carboxyl groups to form a CNT single layer or
25 single layer pattern on the surface of substrate by amidation reaction between the

amine group and the carboxyl group; (b) reacting the CNT single layer or single layer pattern with a diamine type organic compounds to modify the CNT single layer with an organic amine group and reacting the organic amine with the CNT having exposed carboxyl groups to laminate a CNT layer thereon; and (c) repeating the step (b) n times
5 to form CNT layers and organic amine groups laminated alternately for n times, thereby forming a high density CNT film or pattern having exposed carboxyl groups.

The present invention also provides a high density CNT film or pattern which is prepared by the above-described method and has a carboxyl group exposed on its surface.

10 The present invention also provides a CNT-biochip comprising a bio-receptor fixed to the carboxyl group exposed on the CNT film or pattern by chemical or physicochemical bond, in which the bio-receptors have a functional group capable of binding to the carboxyl group and a method for fabricating the same.

Also, the present invention provides a method for producing a high density
15 CNT film or pattern having a chemical functional group selected from the group consisting of amine group, aldehyde group, hydroxyl group, thiol group and halogen, exposed on its surface, which comprises the steps of (a) reacting a substrate having amine groups exposed on the surface or a substrate having amine groups exposed in a pattern with CNT having exposed carboxyl groups to form a CNT single layer or single
20 layer pattern on the surface of substrate by amidation reaction between the amine group and the carboxyl group; (b) reacting the CNT single layer or single layer pattern with a diamine type organic compound to form an organic amine layer on the CNT single layer and reacting the organic amine with the CNT having exposed carboxyl groups to laminate a CNT layer thereon; (c) repeating the step (b) n times to form CNT
25 layers and organic amine layers laminated alternately for n times, thereby forming a

high density CNT film or pattern having exposed carboxyl groups; and (d) modifying the high density CNT film or pattern having exposed carboxyl groups with a chemical compound having both a functional group capable of binding to the carboxyl group and a chemical functional group selected from the group consisting of amine group, aldehyde group, hydroxyl group, thiol group and halogen.

The present invention also provides a high density CNT film or pattern which is prepared by the above-described method and has a chemical functional group exposed on its surface, in which the chemical functional group is selected from the group consisting of amine group, aldehyde group, hydroxyl group, thiol group and halogen.

The present invention also provides a CNT-biochip comprising a bio-receptor fixed to a chemical functional group, selected from the group consisting of amine group, aldehyde group, hydroxyl group, thiol group and halogen, exposed on the CNT film or pattern by chemical or physicochemical bond, in which the bio-receptor has a functional group capable of binding to the chemical functional group, and a method for fabricating the same.

According to the present invention, the substrate having amino functional groups exposed on its surface can be prepared by treating a substrate with aminoalkyloxysilane, the substrate having the amine groups exposed in a pattern can be prepared by forming a photoresist or organic supra-molecule pattern on a substrate having the exposed amine groups. CNT can be laminated or fixed on such pattern in the vertical or horizontal direction. In the case of a nanopattern of an organic supra-molecule, CNT is preferably fixed in the vertical direction. The substrate having the amine groups exposed in a pattern is prepared by forming a photoresist pattern on a substrate having exposed amine groups using photolithography which is commonly used in the semiconductor process, or by forming a photoresist or organic supra-

molecule pattern on a substrate, followed by treatment with aminoalkyloxysilane.

According to the present invention, the chemical functional group capable of binding to carboxyl group is preferably amine group or hydroxyl group. Also, the bio-receptor can be enzyme substrates, ligands, amino acids, peptides, proteins, DNA, RNA, PNA, lipids, cofactors or carbohydrates, which have carboxyl group, amine group, hydroxyl group, aldehyde group, or thiol group. The target biomaterial can be a substance able to serve as a target by reacting with or binding to the bio-receptor to be detected, including preferably proteins, nucleic acids, antibodies, enzymes, carbohydrates, lipids or other biomolecules derived from living bodies, more preferably DNA or proteins.

According to the present invention, the chemicals having both the functional group capable of binding to carboxyl group and the chemical functional group selected from the group consisting of amine group, aldehyde group, hydroxyl group, thiol group and halogen include $H_2N-R_1-NH_2$, H_2N-R_2-CHO , H_2N-R_3-OH , H_2N-R_4-SH , or H_2N-R_5-X in which R_1 , R_2 , R_3 , R_4 and R_5 are independently a C_{1-20} saturated hydrocarbon, un-saturated hydrocarbon or aromatic organic group and X is halogen element.

Also, the present invention provides a method for detecting a target biomaterial capable of binding to or interacting with a bio-receptor, wherein the method is characterized by using the CNT-biochip according to the present invention.

Also, the present invention provides a CNT-DNA chip using DNA as a bio-receptor and a method for detecting DNA hybridization, wherein the method is characterized by using the CNT-DNA chip.

The term "CNT-biochip" used herein inclusively refers to composites having a bio-receptor chemically or physicochemically bonded to a CNT pattern and can be defined as biochips comprising a bio-receptor attached to a high density CNT pattern

or film by chemical or physicochemical bond (particularly, amide bond).

According to the present invention, the CNT-biochip capable of detecting various types of target biomaterials directly or by an electrochemical or electric signal is fabricated by repeatedly laminating CNT on a solid substrate coated with a chemical functional group (amine group) by chemical bond to prepare a high surface density CNT pattern or film having exposed carboxyl groups and attaching a bio-receptor having a functional group (amine group, hydroxyl group, etc.) capable of chemically reacting with the carboxyl groups to the produced CNT pattern or film.

Meanwhile, in order to attach a bio-receptor without a functional group capable of binding to the carboxyl group, the CNT film or pattern having the exposed carboxyl group is modified with a chemical having both a chemical functional group (amine group, hydroxyl group, etc.) capable of binding to the carboxyl group and a chemical functional group capable of binding to the functional group of the target bio-receptor (amine group, hydroxyl group, thiol group, aldehyde group, etc.). Therefore, nearly all bio-receptors can be chemically or physicochemically attached to the high density CNT film or pattern.

For example, in order to attach a bio-receptor having thiol group, a CNT film or pattern is firstly modified with a chemical having both a chemical functional group capable of binding to the carboxyl group and the thiol functional group (Ex.: $\text{NN}_2\text{-R}_2\text{-SH}$) so that the thiol functional group is exposed on the surface of the CNT film or pattern. Then, a bio-receptor having a thiol group is attached to the CNT film or pattern by S-S bond formation.

According to the present invention, by overcoming the limit of the conventional technologies growing CNT using a catalyst fixed at a predetermined position, it is possible to form a pattern in a desired shape at a desired position. Also, the present

invention has improved the defects involved in the conventional technologies by forming a pattern on a substrate using a polymer or an organic supra-molecule so as to utilize advantage of chemical methods at maximum.

According to the present invention, an electric power source can be connected
5 through at least one conductive nanowire so that charge can be applied to each liquid phase comprising the target biomaterials placed on the CNT or CNT chip, in which the conductive nanowire can be formed as a single atom according to the conventional technology (Kouwenhoven, L., *Science*, 275:1896-97, 1997), by forming a predetermined pattern on a conductive metal and depositing a wire, through which an
10 electric current can flow, by ion implantation or sputtering.

BRIEF DESCRIPTION OF THE DRAWINGS

15 FIG. 1 is a schematic view of the process for preparing a high density CNT film by laminating CNT having exposed carboxyl groups by amidation reaction on a substrate having exposed amine groups.

FIG. 2 shows the process for hybridization of complementary DNA to a CNT-DNA chip prepared by attaching DNA having amine groups to a CNT pattern or film
20 having exposed carboxyl groups.

FIG. 3 shows the process for hybridization of complementary DNA to a CNT-DNA chip prepared by modifying a CNT pattern or film having exposed carboxyl groups with amine groups and attaching DNA having carboxyl group as the terminal group thereto.

FIG. 4 is an XPS spectrum for phosphorous detected on the surface of the CNT pattern or film having DNA chemically bonded.

FIG. 5 shows the result of the hybridization using a DNA chip comprising DNA fixed on a high density CNT pattern, in which (a) shows a fluorescent image of the substrate comprising CNT having exposed carboxyl groups fixed with high density, before binding of DNA, (b) shows the result of the fluorescence detection upon hybridization with complementary DNA, and (c) shows the result of the fluorescence detection upon hybridization with non-complementary DNA.

FIG. 6 shows the result of the hybridization using a DNA chip comprising DNA fixed on a high density CNT film, in which (a) shows a fluorescent image of the high density CNT film, before binding of DNA, and (b) shows the result of the fluorescence detection on the hybridized sample, in which (1) is hybridized with complementary DNA and (2) is hybridized with non-complementary DNA.

FIG. 7 shows the result of the hybridization using a DNA chip comprising DNA fixed on a high density CNT film modified with amine group, in which (a) shows the result of the fluorescence detection upon hybridization with complementary DNA, and (b) shows the result of the fluorescence detection upon hybridization with non-complementary DNA.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

The present invention will hereinafter be described in further detail by examples. However, it is to be understood that these examples can be modified into other various forms, and the scope of the present invention is not intended to be limited to such

examples. Such examples are given to more fully describe the present invention for a person skilled in the art.

Reference will now be made in detail to the preferred embodiment of the present invention, an example of which is illustrated in the accompanying drawings.

5

Example 1: Preparation of CNT having exposed carboxyl groups

The CNT, which can be used in the present invention, is not particularly limited and can be commercially available products or prepared by a conventional method. Pure CNT should be carboxylated at its surface and/or both ends to be used in the present invention.

10

The pristine CNT was refluxed in a nitric acid for 45 hours at 90°C and centrifuged. The residue was washed in distilled water and filtered through a 0.2μm filter. The purified CNT was cut in a sonicator containing an oxidizing acid (a mixture of nitric acid and sulfuric acid) for 16 hours. The cut CNT having exposed carboxyl groups was filtered through a 0.1μm filter to obtain CNT with a predetermined size.

15

Example 2: Preparation of a substrate having exposed amine group

In the present invention, the substrate having exposed amine group was prepared by modifying with aminealkyloxysilane on a substrate such as silicon, glass, melted silica, plastics, PDMS (polydimethylsiloxane). However, commercially available substrates, which had been surface-treated with amine, can also be used.

20

Example 3: Preparation of a high density CNT film having carboxyl group exposed on its surface

The CNT having exposed carboxyl groups, prepared in Example 1, was reacted with the substrate having exposed amine groups, prepared in Example 2 to form a CNT single layer on the substrate by amide bond formation between the carboxyl group and the amine group (FIG. 1(a)).

5 Then, the CNT attached to the substrate by amide bond was reacted with a diamine type organic compound having double amine functional groups while CNT having exposed carboxyl groups was reacted with amine groups at the other side of the diamine type organic compound to form another CNT layer by the formation of amide bond (FIG. 1(b)).

10 Next, the chemical reaction between the CNT having exposed carboxyl groups and the diamine type organic compound was repeated to prepare a high density CNT film comprising the CNT layer and the organic diamine layer laminated alternately for n times and having carboxyl groups exposed on its surface (FIG. 1(c)).

The diamine type organic compound which can be used in the present invention
15 includes compounds having a formula of $\text{HN}_2\text{-R}_1\text{-NH}_2$, in which R_1 is C_{1-20} saturated hydrocarbons, un-saturated hydrocarbons or aromatic organic group.

To accelerate the formation of the above amide bond, HAMDU(*O*-(7-azabenzotriazol-1-yl)-1,3-dimethyl-1,3-dimethylenuronium hexafluorophosphate), DCC(1,3-dicyclohexyl carbodiimide), HAPyU(*O*-(7-azabenzotriazol-1-yl)-1,1:3,3-bis(tetramethylene)uronium hexafluorophosphate), HATU(*O*-(7-azabenzotriazol-1-yl)-
20 1,1:3,3-tetramethyluronium hexafluorophosphate), HBMDU(*O*-(benzotriazol-1-yl)-1,3-dimethyl-1,3-dimethylenuronium hexafluorophosphate), or HBTU(*O*-(benzotriazol-1-yl)-1,1:3,3-tetramethyluronium hexafluorophosphate) is preferably used as a coupling agent, and DIEA(diisopropylethylamine), TMP(2,4,6-trimethylpyridine), or NMI(*N*-
25 methylimidazole) is preferably used as a base.

Also, in the case of using water as solvent, EDC(1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride) is preferably used as a coupling agent, and NHS(*N*-hydroxysuccinimide) or NHSS(*N*-hydroxysulfosuccinimide) is preferably used as a coupling agent (base).

5 In this Example, HATU was used as a coupling agent and DIEA was used as a base. The coupling agent participates in the formation of the amide bond (-CONH-) between the -COOH functional group and the -NH₂ functional group, and the base agent acts to increase the efficiency when the coupling agent forms the amide bond.

10 **Example 4: A substrate having amine groups exposed in a pattern and formation of a high density CNT pattern using the same**

In order to expose amine groups in a pattern on a substrate, 2 types of methods can be used. The first method includes forming a photoresist or organic supra-molecular pattern on a substrate such as silicon, glass, melted silica, plastics, PDMS
15 (polydimethylsiloxane) and fixing aminoalkyloxysilane on the substrate surface using the formed pattern as a mask to expose amine groups in a pattern on the substrate surface. The second method includes treating a substrate surface with aminoalkyloxysilane and forming a photoresist or organic supra-molecular pattern to expose amine groups in a pattern on the substrate surface. A preferred example of
20 aminoalkyloxysilane is aminopropyltriethoxysilane.

Using the substrate having amine groups exposed in a pattern, the process described in Example 3 was repeated to form a high density CNT pattern having carboxyl groups exposed on its surface.

The CNT pattern having exposed carboxyl groups can be modified by chemicals
25 having both a chemical functional group (amine group, hydroxyl group, etc.) capable

of reacting with the carboxyl group and a chemical functional group (amine group, hydroxyl group, thiol group, aldehyde group, etc.) capable of binding to a functional group of a desired bio-receptor. The chemicals which can be used in such modification include $H_2N-R_1-NH_2$, H_2N-R_2-CHO , H_2N-R_3-OH , H_2N-R_4-SH , H_2N-R_5-X and the like, in which R_1 , R_2 , R_3 , R_4 and R_5 are independently a C_{1-20} saturated hydrocarbon, unsaturated hydrocarbon or aromatic organic group and X is halogen element.

Example 5: Fabrication of a DNA chip comprising DNA fixed on a CNT film

A CNT-DNA chip was fabricated by attaching amine groups of DNA to the CNT film having exposed carboxyl groups, prepared in Example 3 (FIG. 2). In this Example, EDC was used as a coupling agent for the formation of the amide bond between the carboxyl group and the amine group and NHS was used as a base agent. Also, in this Example, a CNT-DNA chip was fabricated using oligonucleotide having the following SEQ ID NO: 1 having amine group as the terminal group.

SEQ ID NO 1: 5'-TGT GCC ACC TAC AAG CTG TG-3'

The existence of DNA on the CNT film was confirmed by XPS (X-ray photoelectron spectroscope) spectrum for phosphorus atom considering the fact that all DNAs have phosphate groups (FIG. 4). As shown in FIG. 4, phosphorus was detected in the XPS surface analysis and thus, it was confirmed that DNA was attached to the CNT surface.

In this Example, a CNT-DNA chip was fabricated by attaching carboxyl groups of DNA to the CNT film having amine groups exposed on the surface, prepared in Example 3 (FIG. 3). In this Example, EDC was used as a coupling agent for the formation of the amide bond between the carboxyl group and the amine group and NHS was used as a co-coupling agent. Also, in this Example, a CNT-DNA chip was

fabricated using oligonucleotide having the SEQ ID NO 1 having carboxyl group as the terminal group.

Example 6: Fabrication of a DNA chip comprising DNA fixed on a CNT pattern

5 In this example, a DNA chip was fabricated by attaching amine group of DNA to the CNT pattern having exposed carboxyl groups, prepared in Example 4 (FIG. 2). Alternatively, a DNA chip can be fabricated by modifying the CNT pattern having exposed carboxyl groups, prepared in Example 4, with a diamine type organic compound having amino functional groups at both sides to expose amino functional groups and attaching carboxyl groups of DNA to the amine groups (FIG. 3).
10

Example 7: Analysis of hybridization using a CNT-DNA chip

The DNA chip prepared in Example 6 was placed in a hybridization chamber and a hybridization solution was dropped at where the CNT had been fixed. Then, a
15 cover slide was placed thereon. Here, the hybridization solution was prepared with 32 μl of a solution containing an oligonucleotide of complementary sequence to be a total volume of 40 μl at a final concentration 3XSSC (0.45M NaCl, 0.045M sodium citrate) and 0.3% SDS(sodium dodecyl sulfate). The complementary oligonucleotide sequence was the following SEQ ID NO 2 having FITC (fluorescein isothiocyanate)
20 attached to its end.

SEQ ID NO 2: 5'- CAC AGC TTG TAG GTG GCA CA-3'

The solution was left at 100°C for 2 minutes and centrifuged for 2 minutes at 12000rpm to remove non-specific binding between two oligonucleotide strands. In order to prevent the hybridization solution from being dried in the hybridization
25 chamber, 30 μl of 3XSSC (0.45M NaCl, 0.045M sodium citrate) was placed in each

hollow at both sides of the chamber and the chamber was closed and hybridized for 10 hours at 55°C in a incubator.

The hybridization was detected through a fluorescent image using FITC labeled at the end of the oligonucleotide of the SEQ ID NO 2. The fluorescent image was
5 obtained using ScanArray 5000 (Packard BioScience, BioChip Tecnologies LLC) confocal microscope and the QuantArray Microarray Analysis Software (FIG. 5). In FIG. 5, (a) shows a fluorescent image of the substrate comprising CNT having exposed carboxyl groups fixed thereon with high density, before binding of DNA, (b) shows the result of the fluorescence detection upon hybridization with complementary DNA, and
10 (c) shows the result of the fluorescence detection upon hybridization with non-complementary DNA. It was confirmed that the fluorescence was clear and even when the oligonucleotide having the sequence complementary to the CNT-DNA chip was hybridized (FIG. 5(b)). However, in the CNT pattern without the oligonucleotide fixed thereon (FIG. 5(a)) and in the CNT-DNA chip hybridized with the oligonucleotide
15 having the non-complementary sequence (FIG. 5 (c)), no fluorescence was observed. From these results, it was confirmed that the non-specific reaction almost never occurred.

Also, a hybridization was performed following the process as described above using the CNT-DNA chip prepared in Example 5 (FIG. 6). FIG. 6(a) shows a
20 fluorescent image of the high density CNT film, before binding of DNA, and FIG. 6(b)(1) is the result of the fluorescence detection upon hybridization with complementary DNA and (2) is the result of the fluorescence detection upon hybridization with non-complementary DNA. When the oligonucleotide having the sequence complementary to the oligonucleotide bonded to the CNT film was
25 hybridized, the fluorescence was clear and even ((1) of FIG. 6(b)). However, in the

CNT film without fixed oligonucleotide (FIG. 6(a)) and in the CNT-film hybridized with the oligonucleotide having the sequence non-complementary to the oligonucleotide bonded to the CNT film ((2) of FIG. 6(b)), no fluorescence was observed. From these results, it was confirmed that the non-specific reaction almost
5 never occurred.

In this Example, a hybridization was performed following the process as described above using the CNT-DNA chip prepared in Example 5 (FIG. 7). In FIG. 7, (a) shows the result of the fluorescence detection upon hybridization with complementary DNA and (b) shows the result of the fluorescence detection upon
10 hybridization with non-complementary DNA. As shown in FIG. 7, it was possible to certainly distinguish between the hybridized sample and the non-hybridized sample.

As described above, the present invention provides a high density CNT film produced by repeatedly fixing CNT having carboxyl groups exposed on a substrate
15 having exposed amine groups by amidation reaction and a biochip comprising a bio-receptor attached chemically or physicochemically to a chemical functional group on the surface of the CNT film. Also, the present invention provides a biochip comprising a bio-receptor bonded chemically with a high density CNT pattern produced by laminating CNT having exposed carboxyl groups at a desired position on a substrate.

20 According to the present invention, it is possible to fabricate various types of CNT-biochips by chemically or physicochemically attaching various bio-receptors to a CNT pattern (or film) having exposed carboxyl groups or a CNT pattern (or film) having the exposed functional groups modified by various chemical groups. Also, it is possible to fabricate a CNT-biochip comprising bio-receptors attached evenly with a
25 high density on a surface of a CNT film where chemical functional groups are

abundant and present evenly. Further, the chemical functional groups on the CNT surface can be modified into various functional groups by chemical manipulation.

Particularly, upon fluorescent measurement of DNA hybridization using the CNT-DNA chip according to the present invention, it is possible to reduce unnecessary
5 signals, thereby producing excellent results. The CNT-DNA chip is useful for genotyping, mutation detection, pathogen identification and the like.

While the present invention has been described with reference to the particular illustrative embodiment, it is not to be restricted by the embodiment but only by the appended claims. It is to be appreciated that those skilled in the art can change or
10 modify the embodiment without departing from the scope and spirit of the present invention.